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## ***PIPER*PLANTS AS AN ALTERNATIVE TO THE CONTROL OF *AEDES AEGYPTI* AND *AEDES ALBOPICTUS*: AN INTEGRATIVE REVIEW**

PLANTAS DO GÊNERO *PIPER* COMO ALTERNATIVA PARA O CONTROLE DE *AEDES AEGYPTI* E *AEDES ALBOPICTUS*: UMA REVISÃO INTEGRATIVA

PLANTAS DEL GÉNERO *PIPER* COMO ALTERNATIVA PARA EL CONTROL DE *AEDES AEGYPTI* Y *AEDES ALBOPICTUS*: UNA REVISIÓN INTEGRATIVA

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## **ABSTRACT**

*Aedes aegypti* and *A. albopictus* are the main vectors of severe arboviruses. Plant-derived natural products may represent an important source of secondary metabolites for controlling these vectors. Therefore, we performed an integrative literature review on the effectiveness of *Piper* spp. as natural alternative to the control of *A. aegypti* and *A. albopictus* in experimental assays. We searched the scientific databases available in PubMed, Scopus, and Google Scholar for articles published between 2002 and 2022. A total of 225 articles were preselected; however, 41 studies were evaluated in detail after applying the exclusion and inclusion criteria. We observed the use of 43 different species of *Piper*, of which *Piper aduncum* (n=9, 21%) was the most commonly used. The plant structures most commonly used in bioassays were the leaves (n = 28; 68%), which were processed as crude extracts (n = 23; 56%) to evaluate the larvicidal effect (n = 36; 88%) on *A. aegypti* (n = 40; 97.5%). The best LC<sub>50</sub> (lethal concentration capable of killing 50% of the organism of interest) was observed for the extract of *P. hispidum* leaves against larvae. The secondary metabolites frequently found in natural products derived from *Piper* were: -selinene, -pinene, pelitorin, E-caryophyllene, linalool, and germacrene D. Thus, this review synthesizes knowledge generated over 20 years to confirm the potential of *Piper* in controlling *A. aegypti* and *A. albopictus*. Our data indicate that insecticidal molecules were found in *Piper* plants with great potential to become new commercial products to control populations of these vectors.

## **KEYWORDS**

Arboviruses. *Piper*. *Aedes*. Essential Oil.

## RESUMO

*Aedes aegypti* e *A. albopictus* são os principais vetores de arboviroses para a população humana. Produtos naturais derivados de plantas podem representar uma importante fonte de metabólitos secundários para o controle desses vetores. Portanto, realizamos uma revisão de literatura integrativa sobre a eficácia de *Piper* spp. como alternativa natural para o controle de *A. aegypti* e *A. albopictus* em ensaios experimentais. Pesquisamos artigos publicados entre 2002 e 2022 nas bases de dados científicas disponíveis no PubMed, Scopus e Google Scholar. Foram pré-selecionados 225 artigos; contudo, 41 estudos foram avaliados detalhadamente após aplicação dos critérios de exclusão e inclusão. Observamos o uso de 43 espécies diferentes de *Piper*, das quais *Piper aduncum* (n=9, 21%) foi a mais utilizada. As estruturas vegetais mais utilizadas nos bioensaios foram as folhas (n = 28; 68%), que foram processadas como extratos brutos (n = 23; 56%) para avaliar o efeito larvicida (n = 36; 88%) sobre *A. aegypti* (n = 40; 97,5%). A melhor CL<sub>50</sub> (concentração letal capaz de matar 50% do organismo de interesse) foi observada para o extrato de folhas de *P. hispidum* contra larvas. Os metabólitos secundários frequentemente encontrados em produtos naturais derivados de *Piper* foram: -selineno, -pineno, pelitorina, E-cariofileno, linalol e germacreno D. Portanto, essa revisão sintetiza o conhecimento gerado ao longo de 20 anos para confirmar o potencial de *Piper* no controle de *A. aegypti* e *A. albopictus*. Nossos dados indicam que moléculas inseticidas foram encontradas em *Piper*, com grande potencial de tornarem-se novos produtos comerciais para controle das populações desses vetores.

## PALAVRAS-CHAVE

Arboviroses. *Piper*. *Aedes*. Óleo Essencial.

## RESUMEN

*Aedes aegypti* y *A. albopictus* son los principales vectores de arbovirus para la población humana. Los productos naturales derivados de plantas pueden representar una fuente importante de metabolitos secundarios para el control de estos vectores. Por lo tanto, realizamos una revisión integradora de la literatura sobre la eficacia de *Piper* spp. como alternativa natural para el control de *A. aegypti* y *A. albopictus* en ensayos experimentales. Se buscaron artículos publicados entre 2002 y 2022 en bases de datos científicas disponibles en PubMed, Scopus y Google Scholar. Se preseleccionaron 225 artículos; sin embargo, 41 estudios fueron evaluados en detalle luego de aplicar los criterios de exclusión e inclusión. Observamos el uso de 43 especies diferentes de *Piper*, de las cuales *Piper aduncum* (n=9, 21%) fue la más utilizada. Las estructuras vegetales más utilizadas en los bioensayos fueron las hojas (n = 28; 68%), las cuales fueron procesadas como extractos crudos (n = 23; 56%) para evaluar el efec-

to larvicida (n = 36; 88%) sobre *A. aegypti* (n = 40; 97,5%). La mejor  $CL_{50}$  (concentración letal capaz de matar el 50% del organismo de interés) se observó para el extracto de hoja de *P. hispidum* contra larvas. Los metabolitos secundarios encontrados frecuentemente en los productos naturales derivados de *Piper* fueron: -selineno, -pineno, pelitorina, E-cariofileno, linalool y germacreno D. Por lo tanto, esta revisión sintetiza el conocimiento generado a lo largo de 20 años para confirmar el potencial de *Piper* en el control de *A. aegypti* y *A. albopictus*. Nuestros datos indican que en *Piper* se encontraron moléculas insecticidas, con gran potencial para convertirse en nuevos productos comerciales para controlar poblaciones de estos vectores.

## PALABRAS CLAVE

Arbovirosis; *Piper*. *Aedes*. Óleo esencial.

## 1 INTRODUCTION

Many plant species have been used in traditional medicine for the treatment of various diseases. Among them, the Piperaceae family stands out with approximately two thousand described species, predominantly located in tropical forests (SAMAIN *et al.*, 2010). The predominant genus belonging to the Piperaceae family is *Piper* (OLIVEIRA *et al.*, 2020) because of its great potential for medicinal purposes (SILVA *et al.*, 2017; OLIVEIRA *et al.*, 2020). The most relevant biological activities associated with *Piper* are anti-inflammatory, antimicrobial, antiparasitic, and anticancer (SALEHI *et al.*, 2019) properties. The secondary metabolites produced by *Piper* plants, such as alkaloids (FACUNDO *et al.*, 2012), amides (KAOU *et al.*, 2010), and flavonoids (FACUNDO *et al.*, 2012; NASCIMENTO *et al.*, 2012), are linked with these biological effects. Therefore, we believe that these plants can possibly be widely used to control mosquitoes that transmit arboviruses (arthropod-borne viral infections) such as *Aedes aegypti* (Linnaeus, 1762) and *A. albopictus* (Skuse, 1894). However, few studies have compiled current information on the subject.

The species, *A. aegypti* and *A. albopictus*, are considered predominantly responsible for the transmission of arboviruses, such as Dengue, Zika, Chikungunya, and Yellow Fever (NÄSLUND *et al.*, 2021). In recent years, the incidence of arboviruses has increased owing to globalisation, the adaptation of vectors to increasing urbanisation, and environmental changes (ROBERT *et al.*, 2020). Thus, there is an urgent need to develop new methods for controlling these vectors, and natural chemical compounds derived from *Piper* may represent an important step in the development of alternative insecticides.

Therefore, the use of *Piper* as a natural alternative for controlling *A. aegypti* and *A. albopictus* over the last 20 years was reviewed in this current study. Information about the main plant structures used, chemical composition of extracts and essential oils, and lethal concentrations of these natural products are discussed.

## 2 METHODS

### 2.1 STUDY TYPE AND DATA COLLECTION

This review systematically synthesises the scientific knowledge produced on a given subject over a defined period (SOUZA *et al.*, 2010). Our guiding question was, “How can natural products derived from *Piper* be effective in experimentally controlling the *Aedes* mosquito?”. A search was performed of the scientific literature available in the NCBI databases, PubMed, Scopus, and Google Scholar, covering studies published in Portuguese and English between 2002 and 2022. To search these databases, we used the descriptors “*Piper*” and “*Aedes*”, in addition to the Boolean operator “and”, as a way of linking the two descriptors and restricting the search to the theme of this review (PIZZANI *et al.*, 2012).

After identifying the articles, the selection stage was performed by removing duplicates (the same work indexed in all three databases). The selected articles proceeded to the eligibility stage, and their abstracts, which were original articles that used *A. aegypti* and/or *A. albopictus* as experimental models, were evaluated based on our inclusion criteria. In our final selection, we applied our exclusion criteria and removed review articles, studies that used only isolated compounds or the association of several natural products, repellency tests (because do not induce mortality), and studies that did not evaluate mortality or mention the plant structure used. The final number of articles in the database was evaluated in detail.

### 2.2 DATA PROCESSING AND ORGANIZATION

The articles obtained after the selection steps were organised in ascending order of year of publication (2002–2022) using Excel® software. The data extracted from the articles were: authors, year, *Piper* species and plant structure used, type of chemical compounds of natural product obtained (extract or oil), species, ontogenic form of *Aedes* used in bioassays (ovicide, larvicide, pupicide, and adulticide), and LC<sub>50</sub> values (µg/mL or mg/cm<sup>2</sup>).

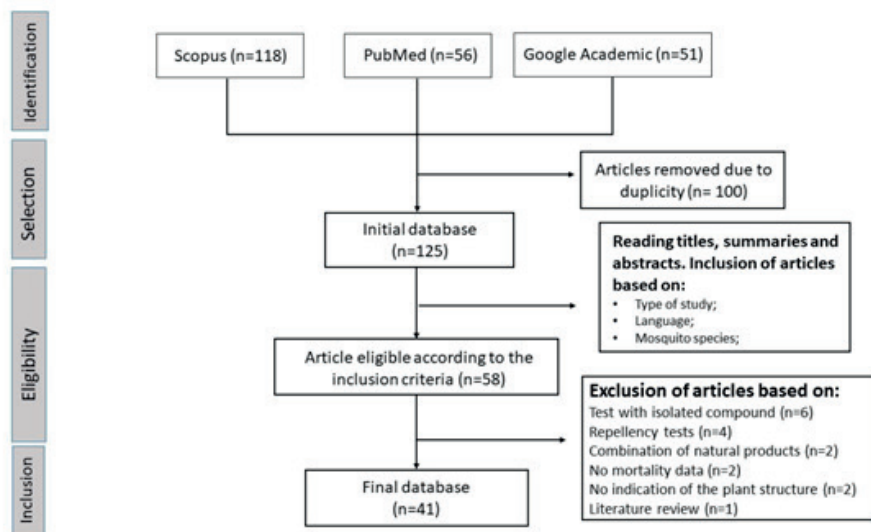
For each piece of information extracted from the articles, the incidence was calculated and organised into graphs and tables. However, it should be noted that some studies evaluated more than one characteristic (e.g., different plant structures or species of *Piper* in the same study); therefore, some graphs exceed 100%.

## 3 RESULTS

In total, 225 scientific research articles were found (Scopus = 118, PubMed = 56, and Google Scholar = 51) using our descriptors (Figure 1). Of these, 100 (approximately 45%) were excluded as duplicates (Figure 1). Of the remaining 125 studies, 58 (approximately 47%) were selected based on the inclusion criteria (Figure 1). In the final selection stage, 17 studies were excluded to our exclusion

criteria. Finally, 41/58 (approximately 71%) studies were evaluated in detail (Figure 1). Information extracted from the selected studies is presented in Table 1.

**Figure 1.** Flowchart of the analysis strategies used in this integrative literature review, which covering studies published between 2002 and 2022 that used *Piper* plants in experimental assays with *Aedes aegypti* and *A. albopictus*.



Of the selected articles, 43 different species of *Piper* were used, as demonstrated in **Table 1**. However, the three most commonly used species were *P. aduncum* (n = 9, 21%), *P. nigrum* (n = 8, 19%), and *P. marginatum* (n = 4, 9%) (Figure 2).

**Table 1.** Data obtained from research articles (2002–2022) that used *Piper* in experimental assays with *Aedes aegypti* and *A. albopictus*.

Species	Plant structure	Product obtained	Species of Aedes	Mosquito phase	Authors
<i>P. aduncum</i>	Leaf	Extract	<i>A. aegypti</i>	Adult	Hidayatulfathi <i>et al.</i> (2004)
<i>P. longum</i>	Fruit, seed and root	Oil	<i>A. aegypti</i>	Adult	Chaiyasit <i>et al.</i> (2006)
<i>P. guineense</i>	Fruit	Extract	<i>A. aegypti</i>	Larva	Oke <i>et al.</i> (2007)
<i>P. nigrum</i>	Fruit	Extract	<i>A. aegypti</i>	Larva	Simas <i>et al.</i> (2007)

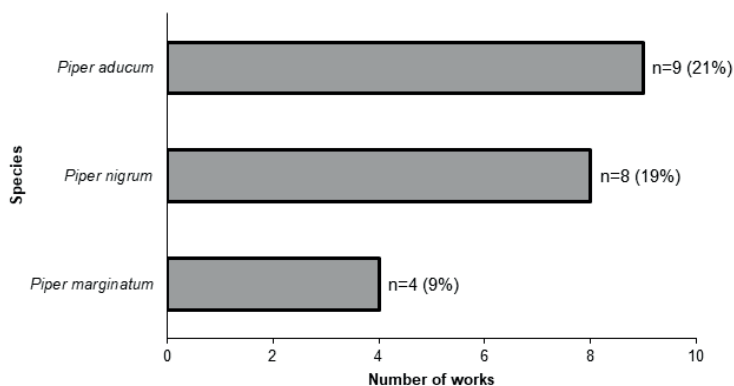
Species	Plant structure	Product obtained	Species of Aedes	Mosquito phase	Authors
<i>P. gaudichaudianum</i> , <i>P. humaytanum</i> <i>P. permucronatum</i> <i>P. hostmannianum</i>	Leaf	Oil	<i>A. aegypti</i>	Larva	Morais <i>et al.</i> (2007)
<i>P. marginatum</i>	Leaf, stalk, and inflo- rescence	Oil	<i>A. aegypti</i>	Larva	Autran <i>et al.</i> (2009)
<i>P. aduncum</i> <i>P. marginalum</i> <i>P. nigrum</i>	Fruit, leaf, and seed	Oil	<i>A. aegypti</i>	Larva	Costa <i>et al.</i> (2010)
<i>P. aduncum</i>	Leaf	Oil	<i>A. aegypti</i> and <i>A. albo-</i> <i>pictus</i>	Adult	Misni <i>et al.</i> (2011)
<i>P. nigera</i>	Seed	Extract	<i>A. aegypti</i>	Adult	Nawaz <i>et al.</i> (2011)
<i>P. nigrum</i>	Seed	Extract	<i>A. aegypti</i>	Larva	Briones <i>et al.</i> (2012)
<i>P. nigrum</i>	Fruit	Extract	<i>A. aegypti</i>	Larva	Grzybowski <i>et al.</i> (2013)
<i>P. ovatum</i>	Leaf, root, and stalk	Extract	<i>A. aegypti</i>	Larva	Kanis <i>et al.</i> (2013)
<i>P. nigrum</i>	Seed	Extract	<i>A. aegypti</i>	Larva	Gulzar <i>et al.</i> (2013)
<i>P. aduncum</i>	Leaf	Oil	<i>A. aegypti</i>	Larva	Oliveira <i>et al.</i> (2013)
<i>P. klotzschianum</i>	Root, stalk, leaf, and seed	Oil	<i>A. aegypti</i>	Larva	Nascimento <i>et al.</i> (2013)
<i>P. nigrum</i>	Fruit	Extract	<i>A. aegypti</i>	Larva	Santiago <i>et al.</i> (2015)
<i>P. arboreum</i> <i>P. marginatum</i> <i>P. aduncum</i>	Leaf	Oil	<i>A. aegypti</i>	Larva	Santana <i>et al.</i> (2015)
<i>P. nigrum</i>	Leaf	Extract	<i>A. aegypti</i>	Larva	Lija-Escaline <i>et al.</i> (2015)

Species	Plant structure	Product obtained	Species of Aedes	Mosquito phase	Authors
<i>P. augustum</i> <i>P. corrugatum</i> <i>P. curtispicum</i> <i>P. darienense</i> <i>P. grande</i> <i>P. hispidum</i> <i>P. jacquemontianum</i> <i>P. longispicum</i> <i>P. multiplinervium</i> <i>P. reticulatum</i> <i>P. trigonum</i>	Leaf	Oil	<i>A. aegypti</i>	Larva	Santana <i>et al.</i> (2016)
<i>P. nigrum</i>	Seed	Extract	<i>A. aegypti</i>	Larva	Custódio <i>et al.</i> (2016)
<i>P. betle</i>	Leaf	Oil	<i>A. aegypti</i>	Adult	Vasantha-Srinivasan <i>et al.</i> (2017)
<i>P. hispidum</i> <i>P. aduncum</i>	Leaf	Extract	<i>A. aegypti</i>	Larva	Porto <i>et al.</i> (2017)
<i>P. betle</i>	Leaf	Oil	<i>A. aegypti</i>	Larva	Vasantha-Srinivasan <i>et al.</i> (2018)
<i>P. ovatum</i>	Root	Extract	<i>A. aegypti</i>	Larva	Kanis <i>et al.</i> (2018)
<i>P. ribesioides</i>	Branch	Extract	<i>A. aegypti</i>	Larva	Kumrungsee <i>et al.</i> (2018)
<i>P. betle</i>	Leaf	Oil	<i>A. aegypti</i>	Egg, larva, and adult	Martianasari; Hamid (2019)
<i>P. aduncum</i>	Inflorescence	Oil	<i>A. aegypti</i>	Larva	Scalvenzi <i>et al.</i> (2019)
<i>P. crocatum</i>	Leaf	Oil	<i>A. aegypti</i>	Larva	Setiawan <i>et al.</i> (2019)
<i>P. klotzschianum</i> <i>P. hispidum</i> <i>P. arboreum</i>	Leaf and stalk	Oil	<i>A. aegypti</i>	Larva	Lima <i>et al.</i> (2019)
<i>P. aduncum</i> <i>P. hispidinervum</i>	Leaf	Oil	<i>A. aegypti</i>	Larva	Silva <i>et al.</i> (2019)

Species	Plant structure	Product obtained	Species of Aedes	Mosquito phase	Authors
<i>P. alatipetiolatum</i>	Leaf	Oil	<i>A. aegypti</i>	Egg, larva, and pupa	Oliveira <i>et al.</i> (2020)
<i>P. longum</i>	Leaf	Extract	<i>A. aegypti</i>	Egg and larva	Dey <i>et al.</i> (2020)
<i>P. corcovadensis</i>	Root	Extract	<i>A. aegypti</i>	Larva	Fernandez <i>et al.</i> (2020)
<i>P. dilatatum</i> <i>P. hostmannianum</i>	Leaf	Oil	<i>A. aegypti</i>	Larva	Albuquerque <i>et al.</i> (2020)
<i>P. capitarianum</i>	Leaf, stalk, and inflorescence	Oil	<i>A. aegypti</i> and <i>A. albopictus</i>	Larva	França <i>et al.</i> (2021)
<i>P. longum</i>	Leaf	Extract	<i>A. aegypti</i>	Larva	Priya <i>et al.</i> (2021)
<i>P. sarmentosum</i>	Leaf	Extract	<i>A. aegypti</i>	Larva	Othman <i>et al.</i> (2021)
<i>P. umbellatum</i>	Leaf	Extract	<i>A. aegypti</i>	Larva	Oliveira <i>et al.</i> (2021)
<i>P. kadsura</i>	Leaf and stalk	Extract	<i>A. albopictus</i>	Larva	Seo <i>et al.</i> (2021)
<i>P. aduncum</i> <i>P. marginatum</i> <i>P. gaudichaudianum</i> <i>P. crassinervium</i> <i>P. arboreum</i> <i>P. hemmendorffii</i> <i>P. cernuum</i> <i>P. lucaenum</i> <i>P. lindbergii</i> <i>P. amalago</i>	Leaf	Oil	<i>A. aegypti</i>	Larva	Pereira-Filho <i>et al.</i> (2021)
<i>P. purusanum</i>	Leaf	Oil	<i>A. aegypti</i> and <i>A. albopictus</i>	Egg and larva	Oliveira <i>et al.</i> (2022)



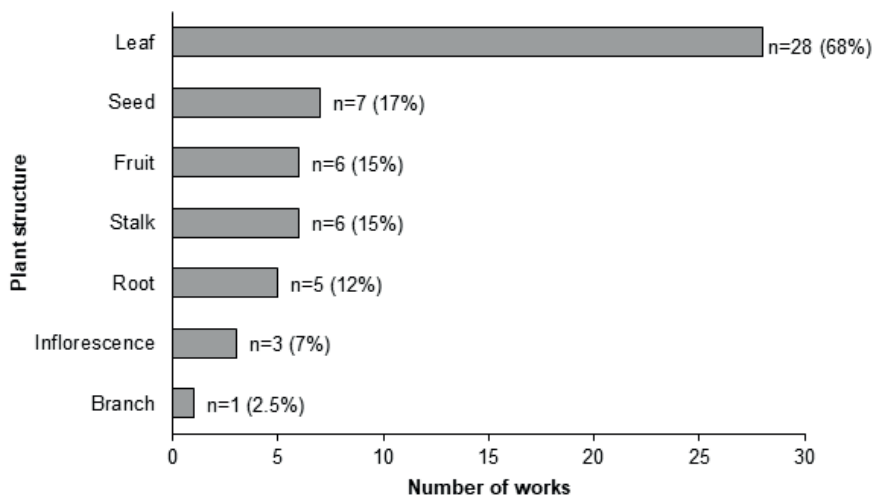
**Figure 2.** The main *Piper* species used in the research (2002–2022) to control *Aedes aegypti* and *A. albopictus*, which were selected from Scopus, PubMed, and Google Scholar databases.



Source: Research data

The plant structures of *Piper* most commonly used were the leaves ( $n = 28$ ; 68%), followed by seeds ( $n = 7$ ; 17%), fruit ( $n = 6$ ; 15%), stalk ( $n = 6$ ; 15%), roots ( $n = 5$ ; 12%), inflorescence ( $n = 3$ ; 7%), and branches ( $n = 1$ ; 2.5%) (Figure 3). Approximately 56% ( $n = 23$ ) of the studies used *Piper* structures to extract crude extracts and 44% ( $n = 18$ ) used *Piper* structures to extract essential oils.

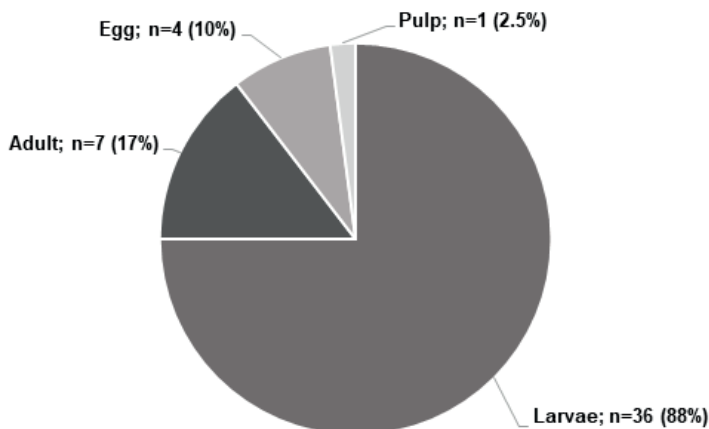
**Figure 3.** Plant structures of *Piper* used in studies (2002–2022) to control *Aedes aegypti* and *A. albopictus*, which were selected from Scopus, PubMed, and Google Scholar databases. The values in graph can exceed 100% because the same study may have included more than one plant structure.



Source: Research data

In the articles research, *A. aegypti* was the most commonly used biological model (n = 40/41; 97.5%), followed by *A. albopictus* (n = 4/41; 10%). Larvae were the mosquito phase most frequently used in the bioassays (n = 36/41; 88%), followed by adults (n = 7/41; 17%), eggs (n = 4/41; 10%), and pupae (n = 1/41; 2.5%) (Figure 4).

**Figure 4.** Ontogenic form most often used in studies (2002–2022) to control *A. aegypti* and *A. albopictus*, which were selected from Scopus, PubMed, and Google Scholar databases. The graph may exceed 100% due to the same work using more than one ontogenic form of mosquito in the bioassays.



Source: Research data

The  $LC_{50}$  values were reported in 33 articles (80.5%). For larvae, studies (n = 31/36; 86%) reported values of 0.17–1174.34  $\mu\text{g}/\text{mL}$ . These data were representative of the extracts of *P. hispidum* leaves and *P. ribesoides* branches. For adults,  $LC_{50}$  values were presented in only three of the seven studies (approximately 43%), with a variation of 0.64–139  $\mu\text{g}/\text{mL}$ . These data are representative of the essential oils from *P. betle* leaves and various plant structures of *P. capitarianum*. The  $LC_{50}$  for eggs was mentioned in only one of the four studies (25%) that used essential oils extracted from the leaves of *P. alatipetiolatum*. The  $LC_{50}$  values obtained in this study was 9.33–65.06  $\mu\text{g}/\text{mL}$ . Studies that used bioassays with pupae did not mention or determine  $LC_{50}$  (Table 2).

Complementally, we also evaluated the chemical compounds identified in the selected studies. Chemical analyses were performed in 28 articles (68%). Thus, it was possible to observe a variety of isolated compounds. Of these compounds, the most frequently mentioned were: -selinene, -pinene, pelitorin, E-caryophyllene, linalool, and germacrene D (Table 2).

**Table 2.** A detailed description of the isolated chemical compounds and LC<sub>50</sub> values of extracts and essential oils of *Piper* structures used in insecticidal bioassays against *Aedes aegypti* and *A. albopictus* mosquitoes in studies published between 2002 and 2022.

Species of <i>Piper</i>	LC50	Chemical compound isolated	Authors
<i>P. aduncum</i>	0.20 mg/cm <sup>2</sup>	NR	Hidayatulfathi <i>et al.</i> (2004)
<i>P. guineense</i>	1.7µg/mL	NR	Oke <i>et al.</i> (2007)
<i>P. nigrum</i>	0.98µg/mL	Piperine and piperolein-A	Simas <i>et al.</i> (2007)
<i>P. gaudichaudianum</i>	121µg/mL	Viridiflorol, aromadendrene, b-selinene, caryophyllene oxide, dillapiole, myristicin, and asaricin	Morais <i>et al.</i> (2007)
<i>P. humaytanum</i>	156µg/mL		
<i>P. permucronatum</i>	36µg/mL		
<i>P. hostmannianum</i>	54µg/mL		
<i>P. marginatum</i>	19.9 – 23.8µg/mL	Monoterpenes, (E)-asarone, and patchouli alcohol	Autran <i>et al.</i> (2009)
<i>P. aduncum</i> <i>P. marginalum</i> <i>P. nigrum</i>	<i>Piper aduncum</i> : 30.2µg/mL <i>Piper marginatum</i> : 8.3 µg/mL <i>Piper nigrum</i> : 75.8µg/mL	-pinene, E-caryophyllene, santolitrene, limonene, caryophyllene oxide, sabinene, iso-elemycin, apiol, and -guaiene	Costa <i>et al.</i> (2010)
<i>P. nigrum</i>	127µg/mL	NR	Briones <i>et al.</i> (2012)
<i>P. nigrum</i>	1.84µg/mL	Piperine	Grzybowski <i>et al.</i> (2013)
<i>P. ovatum</i>	2.9-92µg/mL	Piperlongumine and piperine	Kanis <i>et al.</i> (2013)
<i>P. nigrum</i>	35µg/mL	Pipilyasine, Pipzbedine, Pipyaqubine, Pelitorine, Pipericin, and Piperine	Gulzar <i>et al.</i> (2013)
<i>P. aduncum</i>	134.1 – 289.9 µg/mL	Monoterpenes and sesquiterpenes	Oliveira <i>et al.</i> (2013)
<i>P. klotzschianum</i>	10 – 15.43µg/mL	1-Butyl-3,4-methylenedioxybenzene, 2,4,5-trimethoxy-1-propenylbenzene, limonene, and -phellandrene	Nascimento <i>et al.</i> (2013)
<i>P. nigrum</i>	7.12µg/mL	Oleic acid	Santiago <i>et al.</i> (2015)

Species of <i>Piper</i>	LC50	Chemical compound isolated	Authors
<i>P. arboreum</i> <i>P. marginatum</i> <i>P. aduncum</i>	<i>Piper arboreum</i> : 55µg/mL <i>Piper marginatum</i> : 34µg/mL <i>Piper aduncum</i> : 46µg/mL	Germacrene D, bicyclo-germacrene, (E)-methyl-isoeugenol, (E)-anethole, (Z)-methyl-isoeugen, (E)-isocroweacin, apiole, and elemicin	Santana <i>et al.</i> (2015)
<i>P. nigrum</i>	34.97µg/mL	Thymol and $\zeta$ -elemen	Lija-Escaline <i>et al.</i> (2015)
<i>P. augustum</i> <i>P. corrugatum</i> <i>P. curtispicum</i> <i>P. darienense</i> <i>P. grande</i> <i>P. hispidum</i> <i>P. Jacquemontianum</i> <i>P. longispicum</i> <i>P. multiplinervium</i> <i>P. reticulatum</i> <i>P. trigonum</i>	NR	Cembratrienol, -pinene, -pinene, trans-farnesene, p-cymene, dilapiole, linalool, -phellandrene, limonene, -caryophyllene, linalool, -phellandrene, p-cymene, -selinene, - element, -selinene, and germacrene D	Santana <i>et al.</i> (2016)
<i>P. nigrum</i>	3.1-5.3µg/mL	Piperine	Custódio <i>et al.</i> (2016)
<i>P. betle</i>	0.64-0.72µg/mL	NR	Vasantha-Srinivasan <i>et al.</i> (2017)
<i>P. hispidum</i> <i>P. aduncum</i>	<i>P. hispidum</i> : 0.169µg/mL <i>P. aduncum</i> : 0.342µg/mL	NR	Porto <i>et al.</i> (2017)
<i>P. betle</i>	0.63µg/mL	Eudesm-7(11)-en-4-ol, 2H-cyclopentacyclooctene, 4,5,6,7,8,9-hexahydro-1,2,2,3-tetramethyl and 5-hydroxy-8, 8-dimethyl-3,3a,4,5,6,7,8,8b octahydroindene	Vasantha-Srinivasan <i>et al.</i> (2018)
<i>P. ovatum</i>	2.6µg/mL	NR	Kanis <i>et al.</i> (2018)
<i>P. ribesioides</i>	411.89 - 1174.34µg/mL	Piperine	Kumrungsee <i>et al.</i> (2018)

Species of <i>Piper</i>	LC50	Chemical compound isolated	Authors
<i>P. betle</i>	59.8-183µg/mL	NR	Martianasari; Hamid (2019)
<i>P. aduncum</i>	25.7µg/mL	Dilapiol	Scalvenzi <i>et al.</i> (2019)
<i>P. crocatum</i>	NR	Aromatic compounds and terpenes	Setiawan <i>et al.</i> (2019)
<i>P. klotzschianum</i> <i>P. hispidum</i> <i>P. arboreum</i>	<i>P. klotzschianum</i> : 122.4 - 223.1µg/mL <i>P. hispidum</i> : 141.9µg/mL <i>P. arboreum</i> : 187.9µg/mL	Germacrene D, bicyclogermacrene, (E)-caryophyllene, and -3-carene	Lima <i>et al.</i> (2019)
<i>P. alatipetiolatum</i>	Larvicidal assay: 6.37 - 33.741µg/mL Ovicidal assay: 9.33 - 65.06µg/mL	Sesquiterpenes	Oliveira <i>et al.</i> (2020)
<i>P. corcovadensis</i>	4.86µg/mL	Piperovatin	Fernandez <i>et al.</i> (2020)
<i>P. dilatatum</i> <i>P. hostmannianum</i>	<i>P. dilatatum</i> : 97.56µg/mL <i>P. hostmannianum</i> : 96.13µg/mL	NR	Albuquerque <i>et al.</i> (2020)
<i>P. longum</i>	50.81 - 395.51µg/mL	Eugenol, n-Hexadecanoic acid, Phytol, Oleic Acid, 4-Chromanol, 2,4-Di tert butylphenol, Tetradecanoic acid, Phenol, 4-(2-propenyl), cis-Vaccenic acid, Butanoic acid, 9,12-Octadecadienoyl chloride, (Z,Z)	Dey <i>et al.</i> (2020)
<i>P. capitarium</i>	<i>Larvae/A. aegypti</i> : 82.43 - 139.5µg/mL <i>Larvae/A. albopictus</i> : 63.5 - 133.83µg/mL <i>Adults/A. aegypti</i> : 126.25 - 140.36µg/mL <i>Adults/A. albopictus</i> : 124.5 - 139µg/mL	Rans-caryophyllene, -humulene, -myrcene, -selinene and linalool	França <i>et al.</i> (2021)

Species of <i>Piper</i>	LC50	Chemical compound isolated	Authors
<i>P. longum</i>	0.22 – 0.47 µg/mL	Isopropyl myristate, N-hexadecanoic acid, Dodecane, Heneicosane, and Docosane	Priya <i>et al.</i> (2021)
<i>P. sarmentosum</i>	39.04 – 156.10 µg/mL	Phenylpropanoids (Z-isoolemicin and asarone)	Othman <i>et al.</i> (2021)
<i>P. umbellatum</i>	NR	Tannins, flavonoids, and phenolic compounds	Oliveira <i>et al.</i> (2021)
<i>P. kadsura</i>	NR	Pellitorin, Chingchengenamide A, and Piperenone	Seo <i>et al.</i> (2021)
<i>P. purusanum</i>	<i>A. aegypti</i> : 53.41 µg/mL. <i>A. albopictus</i> : 42.62 µg/mL.	-caryophyllene, -humulene, and germacrene D	Oliveira <i>et al.</i> (2022)

NR = analysis not performed. Studies that did not calculate LC50 and did not investigate the chemical compounds present in natural products derived from *Piper* were not included in this table.

## 4 DISCUSSION

Plants of the *Piper* genus have promising biological activities against *Aedes* spp. and in this review, this knowledge was updated to confirm the potential of *Piper* to produce natural compounds to control these vectors.

Our data demonstrated that the three *Piper* species most commonly used in experiments against *A. aegypti* and *A. albopictus* were *P. aduncum*, *P. nigrum*, and *P. marginatum*. Previous studies (TORRES-PELAYO *et al.*, 2016; DURANT-ARCHIBOLD *et al.*, 2018) have indicated that *P. aduncum* is used in several traditional formulations as an astringent, digestive stimulant, diuretic, antimalarial, sedative, and remedy for haemorrhoids, gonorrhoea, diarrhoea, and toothache. Furthermore, Misni *et al.* (2011) reported that the essential oil of *P. aduncum* has the potential to be used as a spray formulation against adults of *A. aegypti* and *A. albopictus*.

*Piper nigrum* (black pepper) is a widely used spice is known for its chemical constituent called piperine (MEGHWAL; GOSWAMI, 2013). Based on studies in cell cultures, vertebrate experimental models, and humans, piperine has been found to have immunomodulatory, antioxidant, anti-asthmatic, anti-cancer, anti-inflammatory, anti-ulcer, and anti-amoebic properties (MEGHWAL; GOSWAMI, 2013). Furthermore, Lija-Escaline *et al.* (2015) demonstrated that the botanical extract of *P. nigrum* showed remarkable larvicidal activity against *A. aegypti*, severely affecting its intestinal cells.

In addition, *P. marginatum* is popularly known as “pimenta-do-mato”, and it has been demonstrated that the essential oil derived by this plant has strong larvicidal activity against *A. aegypti* (AUTRAN *et al.*,

2009) and fungicidal activity against *Fusarium oxysporum* (SANTOS *et al.*, 2011). Therefore, these data may explain the potential of these three species in experimental studies on *A. aegypti* and *A. albopictus*.

The leaves were most commonly used to obtain extracts and essential oils. Indeed, the leaf is the easiest element of the plant to collect, and its removal induces less damage than other structures (e.g., roots and stems). In most plant species, the leaves are available throughout the year. Although crude extracts are simple to produce, in our study, we did not observe substantial differences in their use in bioassays compared to essential oils. Both natural products are widely commercialised in traditional medicine because of their antioxidant and antimicrobial properties (TONGNUANCHAN; BENJAKUL, 2014). In addition, the use of products derived from plants for insecticide purposes is a tradition more than 3000 years old, which still persists today (PAVELA, 2016).

In the bioassays, *A. aegypti* larvae were the most commonly-used experimental models. This species is considered one of the most important vectors of arboviruses worldwide (NÄSLUND *et al.*, 2021) and has received attention from researchers. Furthermore, the larval stage of *A. aegypti* is the most susceptible to adverse effects compared to the pupal and egg stages (PRICE; MILLS, 1988) which explains the large number of studies that have conducted larvicidal assays.

Lethal concentration (LC) values are frequently determined to measure the biological activity of natural products in experimental bioassays. The crude extract from *P. hispidum* leaves and essential oil from *P. betle* leaves had the lowest LC<sub>50</sub> values in experimental assays against larvae and adults of *A. aegypti*, respectively. These natural products are among the most promising of those observed in the current review. However, it is necessary to investigate the LC<sub>50</sub> values of these two products in nontarget organisms to confirm their environmental safety. These nontarget organisms generally have a reference physiological response that represents various trophic levels of the ecosystems (SILVA *et al.*, 2015). Thus, in these bioassays, it is possible to estimate the extent of the environmental and health impacts of synthetic and natural compounds (LINDE-ARIAS *et al.*, 2008; SILVA *et al.*, 2015). However, toxicity studies on products derived from plants are still poorly performed, because persists the general idea that plants and their components are generally natural and safe (FERRAZ *et al.*, 2022).

In this review, -selinene, -pinene, pelitorin, E-caryophyllene, linalool, germacrene D were frequently found. According to Pereira *et al.* (2014), linalool can enhance the larvicidal effect of the essential oil of *Pimenta dioica* leaves against *A. aegypti*, and Miranda *et al.* (2002) demonstrated that pelitorin had an insecticidal effect against *Diatraea saccharalis* (Lepidoptera: Crambidae). These chemical compounds may explain the biological activity of natural products derived from *Piper* plants against *Aedes* spp. and deserve more attention in future studies.

Although we have found a variety of compounds derived from *Piper* that are biologically active against *A. aegypti* and *A. albopictus*, few commercially available products are still derived from them. Part of this problem is related to the low stability of these products in the environment, which promotes rapid degradation and unsatisfactory biological effects (TUREK; STINTZING, 2013). In addition, many plants are difficult to produce on a large scale, and there is insufficient government support to stimulate the production (PAVELA, 2016). Despite these bottlenecks, studies on the biological effects of products derived from *Piper* plants must continue, to promote the discovery of new highly effective molecules against *Aedes* with reduced environmental toxicity.

## 5 CONCLUSION

Our review demonstrates that the *Piper* genus has the potential to control mosquitoes of the species, *A. aegypti* and *A. albopictus*, which are considered the main vectors of arboviruses worldwide. The most commonly used species were *P. aduncum*, *P. nigrum*, and *P. marginatum*. The leaves were most commonly used for the production of extracts and essential oils to perform larvicidal assays against *A. aegypti*. The biological action of *Piper* against *Aedes* may be connected to the chemical compounds present in the extracts and essential oils, such as -selinene, -pinene, pelitorin, E-caryophyllene, linalool, and germacrene D. This review describes the advances over the years in the study of natural products derived from *Piper* species to fight *Aedes* mosquitoes and may inform new prophylaxis for these vectors.

## REFERENCES

- ALBUQUERQUE, J. S. G. *et al.* Atividade larvicida dos óleos essenciais de *Piper dilatatum* e *Piper hostmannianum* (Piperaceae) para o controle de *Aedes aegypti* (Culicidae) em laboratório. **DeC. Foco.**, v. 4, p. 22-28, 2020.
- AUTRAN, E. S. *et al.* Chemical composition, oviposition deterrent and larvicidal activities against *Aedes aegypti* of essential oils from *Piper marginatum* Jacq. (Piperaceae). **Bioresource Technol**, v. 100, n. 7, p. 2284-2288, 2009.
- BRIONES, A. V. *et al.* Effects of aqueous and pelletized admixture of *Piper nigrum* L. on the oviposition behavior of *Aedes aegypti* mosquitoes and its larvicidal-ovicidal activity. **Acta Med Philipp**, v. 46, n. 3, p. 55-58, 2012.
- CHAIYASIT, D. *et al.* Essential oils as potential adulticides against two populations of *Aedes aegypti*, the laboratory and natural field strains, in Chiang Mai province, northern Thailand. **Parasitol Res**, v. 99, p. 715-721, 2006.
- COSTA, J. G. M. *et al.* Composição química e toxicidade de óleos essenciais de espécies de *Piper* frente a larvas de *Aedes aegypti* L. (Diptera: Culicidae). **Lat Am J Pharm**, v. 29, n. 3, p. 463-467, 2010.
- CUSTÓDIO, K. M. *et al.* A biodegradable device for the controlled release of *Piper nigrum* (Piperaceae) standardized extract to control *Aedes aegypti* (Diptera, Culicidae) larvae. **Rev Soc Bras Med Trop**, v. 49, p. 687-692, 2016.



DEY, P. *et al.* Evaluation of larvicidal activity of *Piper longum* leaf against the dengue vector, *Aedes aegypti*, malarial vector, *Anopheles stephensi* and filariasis vector, *Culex quinquefasciatus*. **S Afr J Bot**, v. 132, p. 482-490, 2020.

DURANT-ARCHIBOLD, A. A. *et al.* Ethnomedical uses and pharmacological activities of most prevalent species of genus *Piper* in Panama: A review. **J Ethnopharmacol**, v. 217, p. 63-82, 2018.

FACUNDO, V. A. *et al.* Non substituted B-ring flavonoids and an indole alkaloid from *Piper aleyreanum* (Piperaceae). **Biochem Syst Ecol**, v. 45, p. 206-208, 2012.

FERNANDEZ, C. M. M. *et al.* Larvicidal activity of piperovatine and dichloromethane extract from *Piper corcovadensis* roots against mosquitoes *Aedes aegypti* L. **Bol Latinoam Caribe**, v. 19, n. 1, p. 142-148, 2020.

FERRAZ, C. A. *et al.* Ecotoxicity of plant extracts and essential oils: A review. **Environ Pollut**, v. 292, p. 118319, 2022.

FRANÇA, L. P. *et al.* *Piper capitarianum* essential oil: a promising insecticidal agent for the management of *Aedes aegypti* and *Aedes albopictus*. **Environ Sci Pollut Res**, v. 28, p. 9760-9776, 2021.

GRZYBOWSKI, A. *et al.* Synergistic larvicidal effect and morphological alterations induced by ethanolic extracts of *Annona muricata* and *Piper nigrum* against the dengue fever vector *Aedes aegypti*. **Pest Manag Sci**, v. 69, n. 5, p. 589-601, 2013.

GULZAR, T. *et al.* New constituents from the dried fruit of *Piper nigrum* Linn., and their larvicidal potential against the dengue vector mosquito *Aedes aegypti*. **Phytochem Lett.**, v. 6, n. 2, p. 219-223, 2013.

HIDAYATULFATHI, O. *et al.* Adulticidal activity of some Malaysian plant extracts against *Aedes aegypti* Linnaeus. **Trop Biomed**, v. 21, n. 2, p. 61-67, 2004.

KANIS, L. A. *et al.* *Piper ovatum* (Piperaceae) extract/starch-cellulose films to control *Aedes aegypti* (Diptera: Culicidae) larvae. **Ind Crop Prod**, v. 122, p. 148-155, 2018.

KANIS, L. A. *et al.* Standardized extract of *Piper ovatum* (Piperaceae) to control *Aedes aegypti* larvae (Diptera: Culicidae). **Ind Crop Prod**, v. 50, p. 816-820, 2013.

KAOU, A. M. *et al.* New amide alkaloid from the aerial part of *Piper capense* L.f. (Piperaceae). **Fitoterapia**, v. 81, n. 6, p. 632-635, 2010.

KUMRUNGSEE, N. *et al.* Bioefficacy of *Piper ribesoides* (Piperaceae: Piperales) extracts and isolated piperine against *Aedes aegypti* (Diptera: Culicidae) larvae and their impact on detoxification enzymes. **Biopestic Int**, v. 14, n. 1, p. 25-32, 2018.

LIJA-ESCALINE, J. *et al.* Physiological and biochemical effects of botanical extract from *Piper nigrum* Linn (Piperaceae) against the dengue vector *Aedes aegypti* Liston (Diptera: Culicidae). **Parasitol Res**, v. 114, p. 4239-4249, 2015.

LIMA, R. N. *et al.* Antitumor and *Aedes aegypti* larvicidal activities of essential oils from *Piper klotzschianum*, *P. hispidum*, and *P. arboreum*. **Nat Prod Commun**. v. 14, n. 7, p. 1934578X19863932, 2019.

LINDE-ARIAS, A. R. *et al.* Multibiomarker approach in fish to assess the impact of pollution in a large brazilian river, Paraiba do Sul. **Environ Pollut**, v.156, n. 3, p. 974-979, 2008.

MARTIANASARI, R., HAMID, P. H. Larvicidal, adulticidal, and oviposition-deterrent activity of *Piper betle* L. essential oil to *Aedes aegypti*. **Vet World**, v. 12, n. 3, p. 367, 2019.

MEGHWAL, M., GOSWAMI, T. K. *Piper nigrum* and piperine: an update. **Phytother Res**, v. 27, n. 8, p. 1121-1130, 2013.

MIRANDA, J. E. *et al.* Potencial inseticida do extrato de *Piper tuberculatum* (Piperaceae) sobre *Alabama argillacea* (Huebner, 1818) (Lepidoptera: Noctuidae). **Rev Bras Olea Fib**, v. 6, n. 2, p.557-563, 2002.

MISNI, N. *et al.* The effect of *Piper aduncum* Linn. (Family: Piperaceae) essential oil as aerosol spray against *Aedes aegypti* (L.) and *Aedes albopictus* Skuse. **Trop Biomed**, v. 28, n. 2, p. 249-258, 2011.

MORAIS, S. M. *et al.* Chemical composition and larvicidal activity of essential oils from *Piper* species. **Biochem Syst Ecol**, v. 35, n. 10, p. 670-675, 2007.

NASCIMENTO, J. C. D. *et al.* Occurrence, biological activities and <sup>13</sup>C NMR data of amides from *Piper* (Piperaceae). **Quim Nova**. v. 35, p. 2288-2311, 2012.

NASCIMENTO, J. C. *et al.* Larvicidal activities and chemical composition of essential oils from *Piper klotzschianum* (Kunth) C. DC.(Piperaceae). **Pest Manag Sci**, v. 69, n. 11, p. 1267-1271, 2013.

NÄSLUND, J. *et al.* Emerging mosquito-borne viruses linked to *Aedes aegypti* and *Aedes albopictus*: global status and preventive strategies. **Vector-Borne Zoonot**, v. 21, n. 10, p. 731-746, 2021.

- NAWAZ, R. *et al.* Adulticidal Activity of *Olea vera*, *Linum usitatissimum* and *Piper nigrum* against *Anopheles stephensi* and *Aedes aegypti* under laboratory conditions. **Iran J Arthropod-Bor**, v. 5, n. 2, p. 2, 2011.
- OKE, O. A. *et al.* Toxicity of hexanolic extract of *Piper Guineense* Schum and Thonn (Piperaceae) seed oil to larvae of *Aedes aegypti* (L). **Eur J Scient Res**, v. 18, p. 6-11, 2007.
- OLIVEIRA, A. C. *et al.* Nanoemulsion loaded with volatile oil from *Piper alatipetiolatum* as an alternative agent in the control of *Aedes aegypti*. **Rev Bras Farmacogn**, v. 30, p. 667-77, 2020.
- OLIVEIRA, A. C. *et al.* Essential oil of *Piper purusatum* C. DC (Piperaceae) and its main sesquiterpenes: Biodefensives against malaria and dengue vectors, without lethal effect on non-target aquatic fauna. **Environ Sci Pollut Res**, v. 29, n. 31, p. 47242-47253, 2022.
- OLIVEIRA, A. K. M. *et al.* Efeito larvívica do extrato etanólíco de *Piper umbellatum* sobre o mosquito *Aedes aegypti*. **South Am J Bas Edu Tec Technol**, v. 8, n. 1, p. 84-101, 2021.
- OLIVEIRA, G. L. *et al.* Chemical study and larvívica activity against *Aedes aegypti* of essential oil of *Piper aduncum* L. (Piperaceae). **An Acad Bras Ciênc**, v. 85, p. 1227-1234, 2013.
- OTHMAN, A. N. *et al.* A comparative study on the larvívica effects of *Piper sarmentosum* (Kaduk) leaves extracts against *Aedes aegypti*. **Pertanika J Sci Technol**, v. 29, n. 4, p. 2811 – 2827, 2021.
- PAVELA, R. History, presence and perspective of using plant extracts as commercial botanical insecticides and farm products for protection against insects—a review. **Plant Protect Sci**, v. 52, n. 4, p. 229-241, 2016.
- PEREIRA, A. I. S. *et al.* Atividade antimicrobiana no combate às larvas do mosquito *Aedes aegypti*: homogeneização dos óleos essenciais do linalol e eugenol. **Educ Quím**, v. 25, n. 4, p. 446-449, 2014.
- PEREIRA-FILHO, A. A. *et al.* Larvívica activity of essential oils from *Piper* species against strains of *Aedes aegypti* (Diptera: Culicidae) resistant to pyrethroids. **Front Plant Sci**, v. 12, p. 685864, 2021.
- PIZZANI, L. *et al.* A arte da pesquisa bibliográfica na busca do conhecimento. **Rev Dig Bibl Cienc Inf**, v. 10, n. 2, p. 53-66, 2012.
- PORTO, K. R. *et al.* Screening of plant extracts and fractions on *Aedes aegypti* larvae found in the state of Mato Grosso do Sul (Linnaeus, 1762) (Culicidae). **An Acad Bras Ciênc**. v. 89, p. 895-906, 2017.

PRICE, L. A., MILLS, K. A. The toxicity of phosphine to the immature stages of resistant and susceptible strains of some common stored product beetles, and implications for their control. **J Stored Prod Res**, v. 24, n. 1, p. 51-59, 1988.

PRIYA, N. R. P., JONES R. D. S. Evaluation of chemical composition and larvicidal efficacy of *Piper longum* L. leaf extract against *Aedes aegypti*. **J Adv Scientific Res**, v. 12, n. 04, p. 256-262, 2021.

ROBERT, M. A. *et al.* Climate change and viral emergence: evidence from *Aedes*-borne arboviruses. **Curr Opin Virol**, v. 40, p. 41-47, 2020.

SALEHI, B. *et al.* *Piper* species: A comprehensive review on their phytochemistry, biological activities and applications. **Molecules**, v. 24, n. 7, p. 1364, 2019.

SAMAIN, M. S. *et al.* *Verhuellia* is a segregate lineage in Piperaceae: more evidence from flower, fruit and pollen morphology, anatomy and development. **Ann Bot-London**, v. 105, n. 5, p. 677-688, 2010.

SANTANA, A. I. *et al.* Chemical composition and biological activity of essential oils from different species of *Piper* from Panama. **Planta Med**, v. 82, n. 11/12, p. 986-991, 2016.

SANTANA, H. T. *et al.* Essential oils of leaves of *Piper* species display larvicidal activity against the dengue vector, *Aedes aegypti* (Diptera: Culicidae). **Rev Bras Plantas Med**, v. 17, p. 105-111, 2015.

SANTIAGO, V. S. *et al.* *Aedes aegypti* larvicide from the ethanolic extract of *Piper nigrum* black peppercorns. **Nat Prod Res**, v. 29, n. 5, p. 441-443, 2015.

SANTOS, M. R. A. *et al.* Antifungal activity of *Piper marginatum* L. (Piperaceae) essential oil on in vitro *Fusarium oxysporum* (Schlecht). **Rev Saúde Pesq**, v. 4, n. 1, p. 9-14, 2011.

SCALVENZI, L. *et al.* Larvicidal activity of *Ocimum campechianum*, *Ocotea quixos* and *Piper aduncum* essential oils against *Aedes aegypti*. **Parasite**, v. 26, a. 23, 2019.

SEO, S. *et al.* Larvicidal activities of *Piper kadsura* (Choisy) Ohwi extract and its constituents against *Aedes albopictus*, toxicity to non-target organisms and development of cellulose nanocrystal-stabilized pickering emulsion. **Ind Crop Prod**, v. 162, p. 113270, 2021.

SETIAWAN, B. U. D. I. *et al.* The effect of red betel leaf's essential oil (*Piper crocatum* Ruiz & Pav.) against third instar *Aedes aegypti* larvae. **P J M H S**, v. 13, n. 4, p. 1162-1165, 2019.

SILVA, J. K. *et al.* Essential oils from neotropical *Piper* species and their biological activities. **Int J Mol Sci**, v. 18, n. 12, p. 2571, 2017.

SILVA, J. D. S. *et al.* Princípios bioéticos aplicados aos estudos ecotoxicológicos aquáticos. **Rev. Bioét**, v. 23, n. 2, p. 409-418, 2015.

SILVA, L. S. *et al.* Encapsulation of *Piper aduncum* and *Piper hispidinervum* essential oils in gelatin nanoparticles: a possible sustainable control tool of *Aedes aegypti*, *Tetranychus urticae* and *Cerataphis lataniae*. **J Sci Food Agr**, v. 99, n. 2, p. 685-695, 2019.

SIMAS, N. K. *et al.* Potential use of *Piper nigrum* ethanol extract against pyrethroid-resistant *Aedes aegypti* larvae. **Rev Soc Bras Med Trop**, v. 40, p. 405-407, 2007.

SOUZA, M. T. D. *et al.* Revisão integrativa: o que é e como fazer. **Einstein (São Paulo)**, v. 8, p. 102-106, 2010.

TONGNUANCHAN, P., BENJAKUL, S. Essential oils: extraction, bioactivities, and their uses for food preservation. **J Food Sci**, v. 79, n. 7, p. R1231-R1249, 2014.

TORRES-PELAYO, V. R. *et al.* A phytochemical and ethnopharmacological review of the genus *Piper*: as a potent bio-insecticide. **Res J Biol**, v. 4, n. 2, p. 45-51, 2016.

TUREK, C.; STINTZING, F. C. Stability of essential oils: a review. **Compr Rev Food Sci Food Saf**, v. 12, n.1, p. 40-53, 2013.

VASANTHA-SRINIVASAN, P. *et al.* Toxicological effects of chemical constituents from *Piper* against the environmental burden *Aedes aegypti* Liston and their impact on non-target toxicity evaluation against biomonitoring aquatic insects. **Environ Sci Pollut Res**, v. 25, p. 10434-10446, 2018.

VASANTHA-SRINIVASAN, P. *et al.* Comparative analysis of mosquito (,plant derived essential oil derived from *Piper betle* L. **Ecotox Environ Safe**, v. 139, p. 439-446, 2017.

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